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2 distribution, survivorship, and aggregate abundance of fall run Chinook salmon. 3 Andrew Olaf Shelton¹ 4 William H. Satterthwaite² 5 Eric J. Ward¹ 6 Blake E. Feist¹ 7 Brian Burke³ 8 9 ¹Conservation Biology Division, Northwest Fisheries Science Center, National Marine 10 11 Fisheries Service, National Oceanic and Atmospheric Administration, 2725 Montlake 12 Blvd. E, Seattle, WA 98112, U.S.A. 13 ² Fisheries Ecology Division, Southwest Fisheries Science Center, National Marine 14 Fisheries Service, National Oceanic and Atmospheric Administration, 110 McAllister 15 16 Way, Santa Cruz, CA 95060, USA 17 ³ Fish Ecology Division, Northwest Fisheries Science Center, National Marine Fisheries 18 19 Service, National Oceanic and Atmospheric Administration, 2725 Montlake Blvd. E, 20 Seattle, WA 98112, U.S.A. 21 22 23

Using hierarchical models to estimate stock-specific and seasonal variation in ocean

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- 27 Supplement S1: Supplemental Tables and Figures
- 28 Supplement S2: Detailed methods
- 29 Supplement S3: Smolt outmigrant data description

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30 Abstract

Ocean fisheries often target and catch aggregations comprised of multiple populations or 31 32 groups of a given species. Chinook salmon originating from rivers throughout the west 33 coast of North America support mixed-stock ocean fisheries and other ecosystem 34 components, notably as prey for marine mammals. We construct the first coastwide statespace model for fall Chinook salmon tagged fish released from California to British 35 36 Columbia between 1977 and 1990 to estimate of seasonal ocean distribution along the 37 west coast of North America. We incorporate recoveries from multiple ocean fisheries and allow for regional variation in fisheries vulnerability and maturation. We show that 38 39 Chinook salmon ocean distribution depends strongly on region of origin and varies 40 seasonally while survival showed regionally varying temporal patterns. Simulations 41 incorporating juvenile production data provide proportional stock composition in 42 different ocean regions and the first coastwide projections of Chinook salmon aggregate 43 abundance. Our model provides an extendable framework that can be applied to 44 understand drivers of Chinook salmon biology (e.g. climate effects on ocean distribution) 45 and management effects (e.g. consequences of juvenile production changes).

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46 Introduction

47 Migratory species present unique challenges for conservationists and managers. A 48 diversity of taxa from insects through mammals occupy and migrate across vast areas of 49 the Earth's surface (Martin et al. 2007; Block et al. 2011), and the movements of many 50 marine fish, marine mammal, and sea turtle species pose challenges for sustainable 51 management as multiple regulatory bodies must collaborate on fishing and management. 52 Population structure, where individuals in a given area consist of multiple, distinct 53 groups, may further complicate marine management. Particular populations, sub-54 populations, or life-history types within a single population often co-occur (e.g. Schindler 55 et al. 2010; Teel et al. 2015; Satterthwaite and Carlson 2015), but the contribution of each 56 group to the aggregate abundance in migratory species may vary spatially and temporally 57 and therefore the importance of a given component in one region often differs from the 58 same component in another. Portfolio theory (Markowitz 1952; Koellner and Schmitz 59 2006) has shown that population complexes with a diverse set of contributing groups will result in reduced variation in aggregate abundance (Hilborn et al. 2003; Schindler et al. 60 61 2010, 2015). Most applications of portfolio theory to natural systems have emphasized 62 the temporal attributes of aggregate abundance, showing how increased diversity among 63 components (Moore et al. 2010; Thorson et al. 2014; Satterthwaite and Carlson 2015) or 64 life-history diversity (Hilborn et al. 2003; Schindler et al. 2010; Greene et al. 2010) lead 65 to resilience and stability in aggregate.

For migratory species, it is important to recognize that the portfolio framework is
relevant in a spatial as well as temporal context (Griffiths et al. 2014). Movement may
create a shifting mosaic in which the distribution of both the aggregate abundance and the

69 individual contributors to abundance shift in space and time. Furthermore, while fisheries 70 are often focused on maintaining robust aggregate abundances over the long term, 71 conservation decisions are often focused on avoiding low abundance for component 72 populations or sub-populations. Management actions to protect or conserve these less 73 productive stocks is generally referred to as 'weak stock' management. Thus, conflict 74 between managing aggregate abundance and the protection of a particular population 75 may arise and strategies for spatial and temporal management must consider this conflict. 76 Chinook salmon (Oncorhynchus tshawytscha) are a highly migratory species 77 native to the Pacific coast of North America. In the eastern Pacific, Chinook salmon 78 occur along the continental shelf and into the open ocean, ranging from central California 79 to Alaska (Healey 1991), where they support extensive and economically valuable 80 fisheries (PFMC 2016a; PSC 2016). Chinook salmon also serve important roles in the 81 ecosystem, as prey for both marine predators such as sharks, pinnipeds, and killer whales 82 (Ford 2012; Chasco et al. 2017) and terrestrial predators including birds and bears (Good 83 et al. 2007; Schindler et al. 2013). Chinook salmon inhabiting any given coastal area are 84 comprised of fish from multiple stocks (Healey 1991; Norris et al. 2000; Weitkamp 85 2010). Understanding the spatial and temporal dynamics of Chinook salmon throughout 86 their range is critical, as some populations of the northeast Pacific are depleted, and listed 87 under the US Endangered Species Act (ESA) or Canadian Species At Risk Act (SARA). 88 While Chinook salmon are one of the iconic species of the northeast Pacific and 89 subject to large-scale fisheries and extensive research (Ruckelshaus et al. 2003), their 90 marine spatial distribution and migration patterns are poorly understood. Some stock-91 specific distributions for juvenile Chinook salmon are available from research surveys

92	(Trudel et al. 2009; Tucker et al. 2011; Burke et al. 2013) while others have provided
93	stock-specific estimates of ocean distribution using coded-wire tags (CWT; Norris et al.
94	2000; Nandor et al. 2010; Weitkamp 2010) or genetic stock identification (Winans et al.
95	2001; Bellinger et al. 2015; Satterthwaite et al. 2015). The vast majority of these tagging
96	and sampling programs are a result of decades of intensive marking and tagging of
97	hatchery raised fish (Nandor et al. 2010; Weitkamp 2010; Satterthwaite et al. 2013).
98	Fisheries management models such as those developed and used annually by the Pacific
99	Salmon Commission's Chinook Technical Committee (CTC; e.g., CTC 2015) use
100	information from CWT recoveries in many fisheries along the coast in concert with
101	spawning escapement to provide information about the abundance and status of Chinook
102	salmon stocks from Oregon to Alaska. The CTC's work provides vital fisheries
103	management advice annually, but it does not include information on Chinook salmon
104	stocks from California and does not provide direct estimates of spatial distribution.
105	In the most comprehensive peer-reviewed coastwide study to date, Weitkamp
106	(2010) examined tag recoveries for 93 Chinook salmon stocks in ocean fisheries from
107	California to the Bering Sea to produce the only comprehensive description of inferred
108	stock-specific spatial distributions. While Weitkamp (2010) was groundbreaking in its
109	breadth and scope, it did not account for fishing effort nor changes in seasonal
110	distribution, meaning that estimated distributions may be biased by uneven fishing and
111	fisheries sampling effort in space and time. Because of differences in stock-specific
112	tagging rates, numbers of tag recoveries could not be compared among stocks to infer
113	relative densities. More targeted studies have inferred season-specific local densities from
114	catch per unit effort (CPUE), typically involving fewer stocks, a smaller spatial range,

and only considering one gear type at a time (Norris et al. 2000; Sharma et al. 2013;
Satterthwaite et al. 2013, 2015). Although Newman (1998) developed a state-space
framework for integrating spatial data on tag recoveries into a demographic model
including mortality and movement, there are limited applications of this approach to
empirical datasets aside from a coho salmon (*Oncorhynchus kisutch*) stock in
Washington (Newman 2000).

121 Here we explore the spatio-temporal dynamics of fall-run Chinook salmon 122 occurring along the Pacific coast of North America using an integrated modeling 123 approach. We use tag release and recovery data from ocean harvest along with data on 124 commercial and recreational fishing effort to estimate a spatio-temporal model for fall 125 Chinook salmon. By modeling all stocks simultaneously, our model shares information 126 among stocks in a biologically reasonable way and leverages the fact that fish derived 127 from different rivers swim in the same areas of the coastal ocean to improve estimates of 128 shared processes. We provide estimates of seasonal ocean distribution and abundance for 129 Chinook salmon populations representing the full geographic extent of North American 130 fall Chinook salmon. To our knowledge, this is the first coastwide analysis of seasonal 131 patterns in density that simultaneously accounts for multiple axes of biological variation 132 among Chinook stocks (differences in maturation, variation in ocean distribution, and 133 spatio-temporal variation in early ocean survival), variation in detection probabilities due 134 to fisheries effort and gear type vulnerabilities, and both measurement and process error. After estimating our biological model, we combine estimates of Chinook salmon ocean 135 136 distribution with regional estimates of juvenile Chinook salmon production to generate 137 estimates of the cumulative abundance and distribution of fall Chinook salmon

abundance along the entire west coast of North America on a seasonal basis. This
unifying statistical framework improves our understanding of Chinook salmon biology
and provides a methodology from which it is possible to explore changing ocean
distributions, spatio-temporal variation in mortality, and interactions with other species
and fisheries.

143 144

145 Study Species

METHODS

146 Chinook salmon (Oncorhynchus tshawytscha) are the largest of the northeast Pacific salmon and native populations spawn in rivers along the northern Pacific Ocean 147 148 from northern Japan to Siberia along the Asian coast and central California to Alaska 149 along the North American coast (Healey 1991; Quinn 2005). Although considerable life-150 history diversity exists both across and within watersheds, populations are typically 151 classified based on the season when adults return to their natal rivers to spawn (run 152 timing, generally designated as fall, winter, spring, and summer runs). Adult run timing 153 may be a good predictor of additional aspects of life-history including the timing of major events in the freshwater phase of the lifecycle (Healey 1991). The life history variation in 154 155 run-timing for adults also translates into differences in when juveniles of each run type 156 migrate to the ocean. For example, fall run Chinook salmon juveniles typically emigrate 157 to sea during their first year of life while spring run fish typically spend an extra year in 158 freshwater before emigrating. There can be considerable variation within runs, and 159 variability in run timing appears to have evolved independently many times (Waples et

al. 2004; Moran et al. 2012). Adults typically spend 2-4 years at sea with northern
populations more often maturing at older ages (Myers et al. 1998; Quinn 2005).

162 Considerable loss and degradation of freshwater habitat, along with a desire to 163 supplement harvest, has led to the establishment of numerous hatchery programs 164 coastwide (Naish et al. 2007). Hatchery production now substantially exceeds natural 165 production in many regions (Table S3.1). Many hatchery fish (and a small number of 166 wild fish) are tagged with CWT (Johnson 1990) which contain a numeric identifier 167 unique to each batch of fish, providing information on stock-of-origin, time of release, 168 and other details about a hatchery release group or wild fish collection event.

169

170 **Data**

171 Although there are multiple Chinook salmon run types on the west coast of North 172 America, fall Chinook salmon are the most dominant and data rich (both in terms of 173 population size and tagging programs). Thus, we restricted our analysis to developing 174 models of fall Chinook salmon ocean distribution, with the idea that these general 175 methods are extendable and applicable to other life-history types and species. We used 176 three sources of data to estimate ocean abundance and distribution. First, we extracted 177 information on CWT releases from the Regional Mark Information System (RMIS; 178 http://www.rmis.org/rmis_login.php?action=Login&system=cwt). We extracted 179 information from tagged releases from 43 major hatcheries spanning central California to 180 Vancouver Island British Columbia and representing fish released between 1978 and 181 1991 (from brood years 1977 to 1990). Fall Chinook salmon are rare in rivers north of 182 British Columbia, where the majority of Chinook salmon runs are spring run. The range

183 of years analyzed was constrained primarily by the availability of fishing effort data (see 184 below) and the high intensity of fishing effort during this period. If the model failed for 185 vears with substantial fishing effort, it would likely fail for more recent years which have 186 seen coastwide declines in Chinook salmon fisheries. For central Oregon to Canada, we 187 selected hatcheries based on their previous identification as major hatcheries associated with indicator stocks by the Pacific Salmon Commission (CTC 2015). Major hatcheries 188 189 from southern Oregon and California were selected based on the indicator stocks used by 190 the Pacific Fishery Management Council (PFMC 2016b). Table S1.1 presents a complete 191 list of hatcheries included in the analysis.

192 For this set of years and hatcheries, we identified 2,196 unique CWT tag codes to 193 include, representing approximately 83 million CWT fish released during the study 194 period (see Table S1.6 for a complete list). This list of tag codes excludes releases where 195 comments indicated major problems with the release (e.g. high disease prevalence). We 196 then aggregated tag codes released by individual hatcheries, brood year, brood stock, 197 release year, and release season. For hatcheries that released fall Chinook salmon at 198 multiple points during the year (i.e., they release both fingerling and yearling Chinook 199 salmon), we categorized tag release into two groups based on season of release. This 200 consolidation resulted in 454 unique hatchery-brood year-release season combinations 201 (see Table S1.1), each of which we refer to as a "release" in subsequent sections. 202 Second, we compiled recovery information for each identified tag code from 203 RMIS. We noted the recovery date, location code, and port at which the fish were 204 sampled. As each tag recovery in the RMIS database has an associated expansion that

aims to correct for the proportion of the catch sampled, we used the expanded number

206 reported for each of the tag codes in the RMIS database. Using the expanded number 207 helps account for temporal and spatial variation in the sampling intensity of the fisheries 208 catch. For marine recoveries, we assigned each recovery to the fishing gear type used, to 209 one of 17 coastal regions (Fig. 1), and to one of four seasons (spring: April-May; 210 summer: June-July; fall: August-October; winter: November-March). Ocean recovery 211 areas were derived largely from those used by Weitkamp (2010). The division of seasons 212 was informed by both the biology of fall Chinook salmon (they enter their natal rivers to 213 begin their spawning migration in the fall) and practical considerations (there is much 214 less information about the spatial distribution of fish in the winter due to reduced salmon 215 fishing effort; see Fig. S1.1-S1.4). We only include recovery information from the three 216 fishing gear types for which we have effort information (see below). In total, this 217 included an estimated 527,711 ocean recoveries for the focal release groups. In addition 218 to the ocean recoveries, we use freshwater recoveries (both from river fisheries and 219 escapement to hatcheries and natural spawning areas) reported in RMIS to parameterize 220 some model components (see *Observation Model* below). As illustrative examples, we 221 provide recovery data from the commercial troll fishery for two releases (Fig. 2). 222 Third, we compiled data on commercial and recreational fishing effort from the 223 United States and Canadian government sources. For commercial troll, treaty troll, and 224 recreational fisheries along the outer coast of Washington, Oregon, and California, we 225 used the PFMC 'blue book' (http://www.pcouncil.org/salmon/background/document-226 library/historical-data-of-ocean-salmon-fisheries/). Recreational effort in Puget Sound, 227 Washington was extracted from published WDFW reports (e.g., WDFW 1979). Alaska 228 troll effort was supplied through a data request to the Alaska Department of Fish and

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229 Game (ADFG) and included both power troll and hand troll gear types. We detail how we 230 combine these two effort types in Supplement S2. We acquired Canadian troll effort 231 through a data request to the Canadian Department of Fisheries and Oceans (DFO) for 232 1982-1995. Earlier years of Canadian troll effort were extracted from official Canadian government data reports (British Columbia Catch Statistics, available at 233 234 http://www.pac.dfo-mpo.gc.ca/stats/comm/ann/index-eng.html). The lack of publically 235 available data describing Canadian commercial troll fishing effort targeting Chinook 236 salmon between 1996 and 2004 limited our analysis to 1979-1995 and to brood years 237 1977-1990. We hope to expand the time-frame in future analyses. Recreational fishing 238 effort data for the study period were also not available from Canada (except for some 239 years in the Strait of Georgia, Fig. S1.4) or from Alaskan waters. We describe how we 240 accounted for these gaps in the model description section below. Complicating matters, 241 each type of effort is reported in different units; recreational effort is reported in units of 242 angler-days in the United States and boat-days in Canada, troll effort is reported in units 243 of boat-days, and treaty troll in units of deliveries (see Supplement S1 and S2 for fishing 244 effort for each gear type)

Troll, treaty troll, and recreational fisheries account for >95% of CWT ocean
recoveries for our release groups. The remaining recoveries were largely from
commercial gillnet and seine fisheries with a few other rare types (e.g. test fisheries).
Many gillnet and seine fisheries incidentally catch Chinook salmon, but some are active
in fisheries in the mouth of the natal river or near the hatchery ("terminal" fisheries).
Since many net fisheries only catch fish from individual sources, they are not a
representative sample of multiple stocks within regions, and including them could affect

model inferences about ocean distribution of Chinook salmon. Therefore, we did not
incorporate data from these sources, but address the implications for these missing
fisheries in the methods and discussion.

255

256 Model

257 To estimate the seasonal abundance and distribution of fall Chinook salmon, we 258 simultaneously model the abundance and distribution of hatchery fall Chinook salmon 259 released into 10 of the 17 ocean regions (Fig. 1) over 14 years (brood years 1977-1990). 260 Our model tracks the abundance of fish from the spring of age 2 (defined as calendar year 261 minus brood year) to fall of age 6, encompassing 19 seasonal time steps. As conventions 262 for describing the age of Chinook salmon are confusing and vary regionally and by run 263 type, we provide a table outlining fall Chinook salmon age classification used here (Table 264 S1.2). Unfortunately, some model components such as fishing mortality vary and are 265 reported by calendar year and season, not model season, and so have appropriate 266 subscripts to reflect this complexity.

267 To generate estimates of the abundance of fall Chinook salmon from distinct 268 regions, we need to quantify at least six core processes: 1) the number fish entering the 269 ocean from natal rivers; 2) the natural mortality of juvenile fish; 3) the natural mortality 270 of adult fish; 4) fishing mortality by age and region; 5) the spatial distribution of fish in 271 the ocean; and 6) the age-specific loss of fish from the ocean due to maturation (salmon 272 leaving the ocean and returning to their natal streams to spawn). We use a state-space 273 framework which separates the biological processes (fish moving, dying from natural 274 causes or fisheries, etc.) from what we observe about these fish populations (generally

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276 populations in locations and areas which may have no observations or missing data. 277 Using parameter estimates from the model and estimates of the number of 278 juvenile fall Chinook salmon, we make projections of the number of Chinook salmon in 279 each ocean region for fish originating from different regions and by age class (see 280 Projected ocean distribution of fall Chinook salmon). Due to the complexity of the model 281 we outline the process and observation models briefly in the main text and highlight 282 model components that are novel to this work. We present a comprehensive model 283 description in online Supplement S2. We provide a full list of parameters and subscripts 284 used in model description in Table S2.1.

fisheries catches). This allows us to explicitly account for and make inferences about

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286 Process Model

287 We track the number and distribution of each Chinook salmon release for the 288 entirety of its life-cycle. Each release is associated with a particular natal region, brood 289 and release year. Because we use hatchery releases, the initial number of fish in each 290 release group is treated as known without error. We estimate an independent juvenile 291 mortality rate - spanning the period from date of release to season 1 of the model - for each release and denote it ϕ_i for release *i*. During the 19 seasonal time steps of the model 292 293 (subscript a; Table S1.2), we model the total abundance of each release coastwide as an unobserved, latent variable, N_{i.a}. In each season, fish are subjected to age-specific natural 294 mortality rate, M_a , and are captured in commercial and recreational fisheries at fishing 295 296 mortality rates, F_{-} (subscripts suppressed, see below) that are determined by the fishing 297 effort in a particular region and the age- and gear-specific vulnerability. Both natural and

fishing mortality are modeled as density-independent processes and modeled as occurring
simultaneously. An important assumption of the model is that fish of the same age in the
same spatial region and season are considered to be equivalently vulnerable to ocean
fisheries occurring in that spatial box and season. We incorporate information on
retention size limits for each year, season, and spatial region (see Table S1.5).
Additionally, we include a process variability term to incorporate additional, unmodeled

304 aspects of fisheries and the environment.

305 The distribution of Chinook salmon among the 17 ocean regions varies among seasons and is estimated within our population dynamic model. We let $\theta_{r,l,s}$ be the 306 proportion of fish from natal region r, present in ocean region l, at the beginning of 307 308 season s, and estimate $\theta_{r,l,s}$ within the model. For a given natal region and season, across 309 all locations the proportions must sum to 1. We assume that fish from the same natal 310 region, but potentially different rivers or hatcheries, have identical ocean distributions in 311 a given season and that ocean distributions within a season are the same across Chinook 312 salmon ages. Weitkamp (2010) suggested that ocean distribution may vary with ocean 313 age (ocean age = recovery year - release year) with very young fish (ocean age = 1) 314 found closer to their natal river mouth than older fish (ocean ages 2 to 5), but with the 315 older age classes being broadly similar in distance from their natal river (their Tables 5. 316 6). As our model starts well into ocean age 1 (using Weitkamp's (2010) age accounting; 317 see Table S1.2) and focuses on modeling older fish – those that are susceptible to ocean 318 fisheries – this model assumption matches available information. Although results from 319 Satterthwaite et al. (2013) indicates modest differences in age-specific distributions of 320 older fish from a single stock, the statistical significance of these differences was not

assessed. Therefore, modifications that allow for age-dependence in ocean distribution
should be an important consideration in future work, but are unlikely to fundamentally
change our conclusions.

324 In the fall, adult Chinook salmon leave the ocean to return to the freshwater 325 spawning grounds. We model spawning as the proportion of fish leaving the ocean in the 326 middle of the fall season. We define a small number of ocean regions near the river 327 mouth from which fish can enter their river to spawn (see Table S1.3). This ensures that 328 fish cannot instantaneously jump thousands of kilometers into their natal river, but does 329 acknowledge that fish from several ocean regions may contribute spawning fish. This 330 assumption differs substantially from other Chinook salmon models which do not 331 explicitly consider spatial distributions (e.g. CTC 2015). We model the proportion of 332 mature fish leaving the ocean as a logistic function of age in years. Again, we 333 acknowledge alternate formulations for leaving the ocean to spawn may be appropriate. 334

335 Observation Model

336 There are few direct, fisheries independent surveys of Chinook salmon in the 337 ocean, but Chinook salmon were caught coastwide across a range of fisheries (but see 338 surveys of very young Chinook salmon; Trudel et al. 2011; Burke et al. 2013). We use spatially explicit recovery data from three fisheries gear types in our analysis 339 340 (commercial troll, recreational hook and line, and commercial treaty troll) to calculate the 341 expected catch of fish from release i, gear g, ocean region l, season s, and calendar year c 342 as a function of the number of age *a* Chinook salmon present and fishing mortality in 343 each region. For winter, spring, and summer seasons (seasons without fish escaping to

freshwater), the catch follows the Baranov catch equation (Baranov 1918; Beverton andHolt 1957)

$$\mu_{i,a,l,g} = \frac{F_{a,s,c,l,g}}{(M_a + \sum_g F_{a,s,c,l,g})} N_{i,a} \theta_{r,l,s} \left\{ 1 - \exp\left[-\left(M_a + \sum_g F_{a,s,c,l,g} \right) \right] \right\}$$
(1)

For the fall season, let $N_{i,a,l,S}$ be the number of Chinook salmon present in the ocean after spawning fish enter the river midway through the season. Then catch for the entire fall season is

$$\mu_{i,a,l,g} = \frac{F_{a,s,c,l,g}}{(M_a + \sum_g F_{a,s,c,l,g})} N_{i,a} \theta_{r,l,s} \left\{ 1 - \exp\left[-0.5 \left(M_a + \sum_g F_{a,s,c,l,g} \right) \right] \right\} + \frac{F_{a,s,c,l,g}}{(M_a + \sum_g F_{a,s,c,l,g})} N_{i,a,l,s} \left\{ 1 - \exp\left[-0.5 \left(M_a + \sum_g F_{a,s,c,l,g} \right) \right] \right\}$$
(2)

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356

We use two likelihoods to connect the estimated catch $(\mu_{i,a,l,g})$ to the observed catch. First, for all year-season-location-gear combinations for which we have either documented fishing effort and catches (all troll fisheries and recreational fisheries in the California, Oregon, Washington, and part of British Columbia) or only observed catches (recreational fisheries in most of Canada and Alaska), we model the probability of observing greater than zero Chinook salmon as a Bernoulli random variable,

$$G_{i,l,g,a} \sim Bernoulli(logit^{-1}[logit(W_{l,s,c,g}\pi_0) + \pi_1 \log(\mu_{i,a,l,g})])$$
(3)

where *G* takes on a value of 1 if the observed catch, *C*, is positive, and a value of 0 otherwise. Here, $W_{l,s,c,g}$ is the fraction of the catch sampled as extracted from the RMIS database (see Table S1.5). The parameters π_0 and π_1 serve to transform the catch to the logit scale and acknowledge that some stocks may be present and caught in fisheries even if the sampling of the catch does not observe them. As sampling effort for CWT has 372

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varied both spatially and through time, we calculated the observed sampling fraction 362 363 from the RMIS database for each tag recovery and aggregated them by season, spatial 364 region, and gear type. We calculated the median value for the sampling fraction among all reported catches in each region and season and set $W_{l,s,c,l}$ to be the median sampling 365 366 fraction. We estimate a single offset, π_0 , which is a proportion bounded between 0 and 1 367 to account for potential non-independence among individual sampled fish in the catch. Finally, we estimate a slope, π_1 , to scale how observation probability increases with 368 369 increases expected catch.

The second component of the likelihood consists of linking the observed catchesto the estimated catches if greater than zero Chinook salmon were observed.

$$C_{i,l,g,a} \sim LogNormal(\log(\mu_{i,a,l,g}), \exp(\sigma_0 + \sigma_1 log(\mu_{i,a,l,g})))$$

if $C_{i,l,g,a} > 0$ (4)

Here observation error term for the dispersion between observed and predicted catch has two parameters (σ_0, σ_1) and allows the observation error to vary with larger values of predicted catch.

As expressed in eq. 3 and 4, our models explicitly acknowledge that our observations of fisheries catches of particular release groups are uncertain (i.e. there is observation error). This contrasts with some models used in salmon management (e.g. CTC 2015) and cohort reconstruction approaches used by other authors (e.g. Coronado and Hilborn 1998; Kilduff et al. 2014) that assume error free observation of catches.

In addition to recoveries from fisheries, we need to account for the Chinook salmon that leave the ocean and return to their natal river or hatchery and complete their life-cycle. Ideally, we would have a likelihood component corresponding to the observed fish in rivers and hatcheries for each release group. Unfortunately, preliminary

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386	examination of the RMIS database revealed notable deficiencies in the freshwater
387	recovery data; we identified some individual tag groups from throughout the study region
388	with many ocean recoveries but zero or near zero freshwater recoveries. Such
389	discrepancies have been noted by other authors (e.g. Baker and Morhardt 2001).
390	Furthermore, we compared freshwater recoveries reported in RMIS with recoveries used
391	in several stock assessments. For example, we were unable to reproduce the results
392	reported for Iron Gate and Trinity River hatcheries in Hankin and Logan (2010), which
393	we know included significant quality control and additions to the data beyond the raw
394	RMIS data. We could not identify which RMIS freshwater recovery data were reliable
395	and which were not and so we elected to incorporate only information about the relative
396	occurrence of different age Chinook salmon in freshwater recoveries, not the actual
397	expanded numbers of total observed freshwater recoveries. We detail these approaches in
398	the online supplement and provide the mean estimated proportion returning at each age
399	for each region in Table S1.4. This aspect of our model is important because by not using
400	information about in river recoveries we rely on catches in ocean fisheries to estimate
401	both spatial distributions and the various parameters that scale overall abundance (e.g.
402	juvenile survivorship and gear specific catchability). As a result, our estimates of
403	parameters that the scale total abundance of Chinook salmon – including most
404	prominently juvenile survival and catchabilities – are difficult to estimate and likely mis-
405	estimated by an unknown factor. However, this factor will apply to all modeled releases
406	and thus does not change the relative order of survivorships among releases. We note that
407	virtually all other estimates of survivorship for salmon based on cohort reconstructions

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face this problem as well (Coronado and Hilborn 1998; Kilduff et al. 2014, 2015; CTC2015).

410 Finally, we added two constraints to penalize biologically unreasonable life-411 histories within the model and help account for our imprecision in freshwater recovery 412 data (see Supplement S2 for details). First, we constrained the model so that on average, 413 between age 1 and 6 greater than 99% of individuals are assumed to leave the ocean by 414 the final model time step. They must either die from natural causes or fisheries or leaving 415 the ocean for freshwater. This ensures the model avoids parameter spaces where there are 416 a large number of old fish present in the ocean (age 7+) and which accords with Chinook 417 biology. Second, we constrained the model so that the total number of fish from a single 418 cohort surviving from release to make a spawning migration to freshwater average (all 419 ages summed) averaged nearly 2%. For both constraints, we allow for substantial 420 variation among releases so individual releases may differ substantively from these 421 average rates (Supplement S2).

422

423 Estimation

We implemented the above state-space model in STAN (Gelman et al. 2015, Carpenter et al. 2016) as implemented in the R statistical language (R Core Team 2016, Stan Development Team 2016). STAN uses a Hamiltonian Monte Carlo (HMC) sampling (Neal 2011; Hoffman and Gelman 2014; see Monnahan et al. 2016 for a description targeted at ecologists). Table S2.1 provides a description of all parameters and prior distributions. STAN estimates the joint posterior distribution of parameters and latent states. For all results reported here we used five chains using a warmup period of 300

431 iterations and 2000 monitoring iterations. We used model diagnostics such as checks for
432 divergent transitions, comparisons among chains (Gelman-Rubin statistics), and posterior
433 predictive checks.

434

435 Analysis of juvenile survival estimates

436 After estimation of the juvenile mortality rate for each of the 454 release groups, 437 we constructed a linear mixed model to understand drivers juvenile variation. We used 438 $\log(\phi_i)$ as the response variable and explained variation in survivorship using number of 439 months between release and model start (*n* month) as a continuous fixed effect and year, 440 origin region, and *n* month nested within origin region as random effects. This model 441 allows fish from different origin regions and fish that spend more time at liberty in the 442 river and ocean to have different juvenile survivorships. Past analyses comparing early 443 mortality among releases have often ignored such attributes (Coronado and Hilborn 1998; 444 Kilduff et al. 2014, 2015).

445

446 Projected ocean distribution of fall Chinook salmon

The above model provides estimates of many parameters that are important for determining the abundance and distribution of Chinook salmon. However, since hatchery releases of Chinook salmon are not tagged with CWT at constant rates, and hatcheryversus natural-origin fish make up substantially different proportions of different stocks, CWT data alone cannot be used to generate estimates of Chinook salmon abundance in the ocean. We use model estimates in conjunction with estimates of out-migrating juvenile Chinook salmon leaving rivers and hatcheries to provide predicted fall Chinook

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454 salmon abundances in space and time. We outline an approach to simulating Chinook 455 salmon using model estimates and show how both the proportional contribution of 456 individual stocks and aggregate abundance change under two illustrative scenarios. 457 Generating predicted fall Chinook salmon distributions require specifying three model components. First, we need estimates of the number of juvenile fall Chinook 458 459 salmon produced by each origin region (including both hatchery and wild). Second, we 460 need to specify the fishing mortality occurring in space and time. Finally, we need to 461 determine scenarios for juvenile mortality.

462 For the first scenario, we compiled available information on juvenile fall Chinook 463 salmon production from hatchery and wild sources for each origin. We present 464 approximate estimates of juvenile production in Table S3.1 and detail the methods and 465 data sources used for each area estimate in Supplement S3. For fishing mortality, we 466 used the median mortality for each area and season estimated across the observed time-467 series (1979-1995). For juvenile mortality, we used a simple assumption: all juvenile fish 468 experience the same mortality regardless of their origin. Together these assumptions are 469 designed to reflect the distribution of fall Chinook salmon in the ocean under typical 470 ocean conditions, not the distribution and abundance in a particular year. We refer to this 471 as the "base" scenario.

For the second scenario, we used the same value of fishing mortality (median) and juvenile mortality (mean, spatially invariant) as the base scenario. But for Chinook salmon production, we reduced hatchery production in Puget Sound (PUSO) by half from 37 to 18.5 million. We then compare the abundance and distribution of Chinook salmon in the ocean under the base and "PUSO hatchery" scenarios to illustrate the consequences

of changing aspects of hatchery production for ocean abundance and distribution. Other assumptions and simulations could be used to generate distributions and abundance under other scenarios but we provide a relatively simple, hypothetical scenario here as an example of the possibilities of this approach.

For both scenarios, we use Monte Carlo methods to sample from the posterior estimates of estimated parameters and simulate abundance and distribution through time. As we have fixed juvenile survival, fishing mortality, and the number of juveniles arising from each region, variation only reflects uncertainty in the spatial distribution and in the parameters associated with spawning. Thus the simulations underestimate the overall uncertainty in abundance and distribution. We also use the average process error for each origin region (Fig. S1.7), which further underestimates uncertainty.

Results

Despite the large amount of data and many latent states and parameters, the model converged and produced reasonable biological estimates for parameters. The effective sample size for all parameters was greater than 1,000 and maximum \hat{R} (a measure of model convergence) was less than 1.01. We focus on two model components in the main text before turning to two simple scenarios to understand the attributes of Chinook salmon ocean aggregations using simulations. We present posterior estimates of model parameters in the online supplement along with figures of other major model components.



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500 We detected strong differences in seasonal ocean distribution among different 501 origin regions for fall Chinook salmon (Fig. 3). A common pattern across stocks was that 502 fish were generally distributed near their origin region. For example, fish originating 503 between California and southern Oregon (SFB, NCA, SOR) remained in United States 504 waters south of the British Columbia border (regions WAC and south) and were observed 505 rarely in Canadian and Alaskan waters in our dataset. Fish from the most northern region, 506 SWVI, were almost never present south of their origin, and were estimated to be almost 507 exclusively in Alaska and Canada. Fish from the Columbia River basin (COL, MCOL, 508 and UPCOL) showed the broadest spatial distribution with significant proportions present 509 in areas from California to Alaska. Virutally all fish estimated to be present in the Salish 510 Sea (PUSO, SGEO) originated there, indicating few Chinook salmon from the outer coast 511 migrate into the Salish Sea.

There was a signature of seasonal distributions in fish from nearly all regions. Fish from a given ocean region tended to be more northerly distributed in summer than in winter-spring, and due to spawning migrations Chinook salmon tend to be located near their region of origin during the fall. Ocean distributions also tend to be spatially less concentrated in the winter-spring. In part, this may reflect the uneven length of the seasons in our model as winter-spring spans seven months (November – May) while summer spans only two (June – July).

519

520 Variation in early mortality

521 Estimates of mortality for each of the 454 release groups, showed wide variation 522 in mortality rates among releases (range of posterior medians for ϕ_i : 1.39 to 3.29; across

release mean = 2.02) corresponding to a range of survivorship of 0.037 to more than
0.248 for the period between release and the start of the model in April of brood year + 2.
As this range includes releases from all origin regions and years, and thus includes
releases with vastly different lengths of time between release and the start of the model
(from 1 to 13 months), such large variation is not unexpected. We further summarized
model estimates of early mortality in two ways.

529 The linear mixed effect model showed a large effect of n month on $\log(\phi_i)$ with 530 longer time periods associated with increase mortality (slope estimate for *n* month: 531 0.002(0.002); mean(SE)), indicating that on average, an increase of one month resulted 532 in an increase of 0.002 in $\log(\phi_i)$. There was strong among region variation in the 533 overall mortality intercept (SD among regions = 0.053) and variation among years (SD =534 (0.033). This result coincides with intuition – fish that spend more time in the river and 535 ocean should have greater mortality- but this result does highlight that many past 536 analyses comparing early mortality among releases have ignored such attributes 537 (Coronado and Hilborn 1998; Kilduff et al. 2014, 2015).

538 Second, to make our results comparable to estimates of previous analyses of 539 Chinook salmon early survivorship (Kilduff et al. 2014), we used estimated model 540 parameters to calculate estimated survivorship to the beginning of fall season, age 2. We 541 estimated survivorship for each release in the absence of fishing which accounted for juvenile mortality, natural mortality, and estimated process error, $S_i = \exp(-\phi_i - \phi_i)$ 542 $\sum_{a=1}^{a=2} [M_a + \omega_{i,a}]$, where $\omega_{i,a}$ is the estimated process variability. For region 543 combinations that had at least three releases in a given year, we calculated a weighted 544 545 mean and weighted standard deviation for each region and converted estimated

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546 survivorship to z-scores (subtracted the among-year mean, divided by the standard 547 deviation among years; Fig. 4). We use z-scores to emphasize that our results should not 548 be used as estimates of absolute survivorship as we do not include release-specific 549 information about freshwater recoveries or recoveries from marine net fisheries. Due to 550 relatively few releases in SOR, COR, and NOR, we combined these three regions to 551 calculate a single mean for the Oregon coast (denoted "OR"). Survivorship between 552 release and age 2 was highly variable among regions and both within and among years. 553 Interestingly, most regions show substantial temporal variation in survivorship, but years 554 of high and low survivorship are not coincident among regions. Survivorship trends from 555 the Columbia River tended to be fairly coincident (Fig. 4b), as did fish from Oregon and 556 California (Fig. 4c), but similar trends among northern stocks were less obvious (Fig. 4a). 557 Overall there was very high variability within regions in some years (e.g. SFB in 1985) 558 indicating strong differences in survivorship among hatchery releases. As our model does 559 not include abundances of CWT recoveries from freshwater or the small number of 560 recoveries from non-target fisheries, we expect these estimated survivorships biased from 561 their true values. However, the patterns of relative survivorship across time and among 562 regions should be correct.

563

564 *Comparing distributions and cumulative abundance*

565 Given the varied production of Chinook salmon among origin regions (Table 566 S3.1), and the distinct ocean distributions of these fish (Fig. 3), it is not surprising that 567 ocean areas vary substantially in proportional compositions and aggregate abundance of 568 Chinook salmon (Fig. 5). In terms of proportional contribution, seasonal variation is

present but not striking. For example, the two southeast Alaska regions (NSEAK and SSEAK) are comprised predominantly of fall Chinook salmon from Canada, Washington, and the Columbia River basin in all seasons, thought the proportion from Columbia basin increases notably from spring to summer. The Salish Sea (regions PUSO and SGEO) are dominated by fish originating in those regions in all seasons, while the Californian ocean regions (MONT, SFB, MEN, and NCA) all have close to or more than 50% of fish present originating from California rivers in all seasons (Fig. 5).

576 While the proportional composition of a given area may be relatively consistent 577 across seasons, the distribution changes for many origin regions simultaneously, resulting 578 in substantial difference among seasons in the cumulative abundance of fall Chinook 579 salmon (Fig. 5). Notably the southern most regions (MONT and SFB) and PUSO have 580 the lowest total abundance in all seasons. In contrast, the northern regions (SSEAK and 581 NSEAK) have relatively low abundance in the spring (Fig. 5a) but the abundance 582 increases markedly during the summer (Fig. 5b) reflecting a northerly shift in 583 distributions of most Chinook salmon stocks (Fig. 3).

The cumulative abundance and distribution of fall Chinook salmon also depend 584 585 strongly on the age range of Chinook salmon considered. For example, the cumulative 586 abundance of fish age 2 and older is substantially different from the distribution of fish 587 age 4 and older (compare Fig. 6e and 6j). Old and large fish are notably more abundant 588 in the northern regions, whereas young and small fish are more available in the southern 589 parts of the range. Note that this change in distribution is not driven by changes in the 590 distribution of fish with age (fish of different ages are modeled as having identical ocean 591 distributions; see *Methods*) but is due instead to strong differences in maturation

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592 probability among origin regions. Fish originating from northern areas tend to mature at 593 older ages (Fig. S1.8; Table S1.4). This is readily apparent in comparing the increasing 594 contribution of WAC and SGEO origin fish to total age 4+ fish relative to age 2+ and the 595 concomitant decline of SFB and COL fish (Fig. 6). 596 Finally, comparisons of the base and PUSO hatchery scenarios reveal how 597 changes in management have ramifications beyond the region of origin. We contrast the 598 projected abundance in total abundance between scenarios for age 3+ during the summer 599 (Fig. 7). From a fisheries perspective, most Chinook salmon are vulnerable to both 600 commercial and recreational fisheries by summer age 3 (model season 10; Table S1.2, 601 Fig. S1.6), and we can consider the changes between the two scenarios as affecting the 602 number of fish potentially available to fisheries in a given region. We clearly show that a 603 reduction of hatchery production by half is predicted to change Chinook salmon 604 abundance most dramatically in Puget Sound - a decline in abundance by nearly one-third 605 between base and PUSO hatchery scenarios – but declines of more than 10% are 606 predicted along the Washington coast (WAC) and southern Canadian regions (SGEO, 607 SWVI, NWVI) as well. Changes to PUSO hatchery production are predicted to have a 608 limited effect on the most southerly and northerly regions. 609

610 **Discussion**

We present a coast-wide model for fall Chinook salmon that simultaneously models populations originating from California to British Columbia and accounts for biological variation among populations and across time. We explicitly account for the fisheries effort and sampling of fisheries that affect the detection of Chinook salmon

615 populations in the ocean. Together, our model provides a joint estimate of salmon spatial 616 distribution, juvenile mortality, and spatio-temporal estimates of fisheries mortality; 617 processes that are typically estimated and discussed separately (e.g. Weitkamp 2010; 618 Kilduff et al. 2014; CTC 2015). By estimating a joint time-series model that includes 619 populations spanning much of the northeastern Pacific, we are able to move beyond 620 comparisons of CPUE of different Chinook salmon stocks derived from CWT 621 (Satterthwaite et al. 2013; Norris et al. 2000) or Genetic Stock Identification (GSI; 622 Bellinger et al. 2015; Satterthwaite et al. 2015) and translate information from fisheries 623 catch into estimates of spatial distribution and total abundance. Importantly, our work 624 explicitly accounts for missing data – locations and times where no one was fishing and 625 therefore no sampling of Chinook salmon occurred – and thus expands on previous 626 examinations of salmon ocean distribution. Our work is a tool has broad application for 627 understanding patterns of spatio-temporal variation among Chinook salmon and other 628 tagged salmonid populations. Additionally, it is a simulation platform for exploring the 629 consequences of biological variation and management decisions on an important marine 630 resource.

We present a step toward understanding the portfolio of Chinook salmon populations contributing to each coastal region in each season. A full exploration of spatial portfolios of Chinook salmon would involve accounting for factors contributing to variation within and covariation among populations and is beyond the scope of this paper. However, basic tenets of portfolio theory do allow us to begin to discuss the implications of the spatial patterns. Broadly, portfolio theory suggest that regions that are more highly reliant on fish originating from one or a few areas would experience more temporal

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638	variability than areas with more contributing populations. In the base scenario, three areas
639	had greater than 50% of their abundance derived from a single region in all seasons: two
640	in California (MONT, SFB) and Puget Sound, Washington (PUSO). Additionally, these
641	three areas are also estimated to have the lowest total Chinook salmon abundance.
642	Together these facts suggest these regions with low stock diversity are likely susceptible
643	to periods of especially low abundance. Indeed, the ocean fishery in California and
644	southern Oregon was recently closed for two consecutive years due to poor production of
645	Sacramento River fall Chinook salmon (Lindley et al. 2009; Carlson and Satterthwaite
646	2011), and our simulation of reduced hatchery production highlights the sensitivity of
647	PUSO to changes in local hatchery production (Fig. 7). In contrast, other regions are
648	composed of fish from diverse sources and have a more balanced contribution (e.g. NOR,
649	SWVI, NWVI, NBC, SSEAK, NSEAK) and would be expected to have more stable
650	portfolios over the long-term. Interestingly, these areas largely correspond to locations
651	with important and historically productive Chinook salmon troll fisheries (Southeast
652	Alaska, west coast Vancouver Island, and Oregon coast).

653 From the perspective of predator populations, increased stock diversity (and 654 stability) may translate to increased growth rates. Anecdotally, piscivorous killer whale 655 populations with higher latitude distributions tend to have higher population growth rates 656 (Ward et al. 2013). When considering portfolios, though, it is important to note that this 657 analysis does not include other Chinook salmon life-history types. Spring-run Chinook 658 salmon are the other major life-history type in the northeastern Pacific; other run-types 659 such as winter run Chinook salmon are confined to California rivers and relatively rare 660 (Quinn 2005), though in some locations summer runs are also present. Spring Chinook

661 become more abundant with increasing latitude as all Chinook originating from Alaskan 662 and northern British Columbian rivers are spring-run. Thus, while this analysis presents a 663 reasonable approximation of the Chinook portfolio in California, it dramatically 664 underestimates both the total number of Chinook and life-history diversity present in 665 British Columbia and Alaskan waters in particular. Further work must be done to 666 incorporate the range of life-histories of Chinook salmon into ocean portfolios. Overall, 667 however, portfolio approaches have clear potential for examining the consequences of 668 aggregate patterns of abundance, how they may affect directed fisheries or incidental 669 catch in non-directed fisheries, ecosystem considerations for species dependent upon 670 aggregate abundance such as killer whales or other marine mammal predators, and how portfolio properties vary in response to management or environmental changes. 671 672 While the outlined model incorporates many important attributes of Chinook 673 biology, it necessarily makes simplifying assumption to accommodate missing or 674 incomplete data and ensure model identifiability. Several major aspects of our estimation 675 model should be the foci of future improvement and research. Most importantly, reliable 676 data pertaining to tag recoveries in the escapement of Chinook to freshwater would 677 greatly improve estimates of both maturation probabilities and ocean survivorship. Such 678 information would have the largest impact on the juvenile survivorship estimates (Fig. 4) 679 and the catchability coefficients (q; see Supplement S2) as they serve to scale the overall 680 abundance of fish available in the ocean for fisheries. However, corralling and verifying 681 such data coast-wide is a major task that is beyond the scope of this project. Other 682 reasonable and important extensions to the model include: a) allowing for age-specific or 683 oceanographic driven changes to seasonal distributions; b) accounting for population

684 specific growth rate and/or temporal variation in growth which would translate into 685 population difference in vulnerability to fishing gear types; c) including the fishing effort 686 data necessary to expand the study time-window to include data from 1996 onward; and 687 d) incorporating information from non-mixed stock fisheries such as terminal gillnet and 688 seine fisheries that are not equally likely to capture fish from different origins. 689 Projections of total Chinook salmon abundance could be substantially improved with 690 improved information about the outmigration of juvenile Chinook salmon from rivers 691 coastwide.

692 Much of the interannual dynamics of ocean mortality for Chinook tends to happen 693 very early after migrating downstream, when size and growth play a large role in survival 694 (Beamish et al. 2004; Duffy and Beauchamp 2011). These complex ecological dynamics 695 vary by season and year and require much more data to describe than we could include in 696 this model. Moreover, disentangling juvenile salmon mortality rates in different habitats 697 (rivers, estuaries, coastal ocean) is an ongoing effort. We applied a simple approach -698 allowing juvenile survival to be independent of adult survival and vary by release - but 699 did not model the full mechanistic processes underlying variation in juvenile mortality. 700 Given the importance of early life-stages on overall population dynamics, connecting this 701 model more closely to early survivorship is likely one of the more crucial aspects to 702 tackle in future work.

An important additional consideration for ocean distribution modeling is understanding how to incorporate information from both physical tagging using older technologies (CWT) and information derived from more recently developed and applied genetic stock identification tools (GSI; Satterthwaite et al. 2014; Bellinger et al. 2015). In

707 practice. GSI data provides information about the proportional contribution of fish from 708 different origins in a given area or catch per unit effort information for different stocks. 709 Using GSI data alone without an accompanying analysis of scales or otoliths lacks 710 information about age-structure. As the age-structure will strongly affect the estimated 711 stock composition of any given ocean region, GSI information alone may provide 712 difficult to interpret patterns (Fig. 6). Overall, however, our predictions (Fig. 5-7) should 713 provide predictions for proportional compositions that can be compared directly to GSI 714 studies. Integrating GSI and CWT data in a single integrated framework is an exciting 715 and important area for future work.

716 Beyond data and model complexity, computational limitations do present a 717 challenge for large models like ours. In theory, there is no constraint upon how many 718 releases can be modeled simultaneously, but the 454 releases modeled here require 719 estimation of nearly 8,600 latent states and incorporate over 228,000 observations for the 720 binomial likelihood (the total number of release-location-season-gear type combinations) 721 and over 17,000 observations for the log-normal component. Expanding the number of 722 releases substantially would require substantially improving computational resources or 723 moving away from full Bayesian estimation toward approximations of the posterior 724 distribution such as Laplace approximations (Rue et al. 2009).

Overall, we provide a framework to integrate information from multiple fall Chinook salmon stocks to simultaneously estimate parameters from a complex population dynamic model. We emphasize the spatio-temporal attributes of the parameters here – particularly estimates of ocean distribution and regional patterns in juvenile survival – and provide illustrative examples of how these estimates can be used to simulate

730 scenarios and that these scenarios may be useful in a variety of management and
731 ecosystem contexts in the future.
732
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740 N00070-16-MP-4C872).

741

742 References

743 Baker, P.F. and Morhardt, E. 2001. Survival of Chinook salmon smolts in the

744 Sacramento-San Joaquin Delta and Pacific Ocean. In: R.L. Brown, ed. Contributions

to the biology of Central Valley salmonids. Fish Bull. 179: Volume 2. California

746 Department of Fish and Game, Sacramento. pp. 163-182

747 Baranov, T.I. 1918. On the question of the biological basis of fisheries. Nauch. Issledov.

748 Iktiol. Inst. Izv. I 1: 81–128.

Beamish, R.J., Mahnken, C., Neville, C.M. 2004. Evidence that reduced early marine
growth is associated with lower marine survival of coho salmon. Trans. Am. Fish.
Soc. 133: 26-33

752 Bellinger, M.R., Banks, M.A., Bates, S.J., Crandall, E.D., Garza, J.C., Sylvia, G., and

753	Lawson, P.W. 2015. Geo-referenced, abundance calibrated ocean distribution of
754	Chinook salmon (Oncorhynchus tshawytscha) stocks across the West Coast of North
755	America. PLoS ONE 10(7): e0131276. doi:10.1371/journal.pone.0131276.
756	Beverton, R.J.H., and Holt, S.J. 1957. On the dynamics of exploited fish populations.
757	Chapman & Hall.
758	Block, B.A., Jonsen, I.D., Jorgensen, S.J., Winship, A.J., Shaffer, S.A., Bograd, S.J.,
759	Hazen, E.L., Foley, D.G., Breed, G.A., Harrison, A.L., Ganong, J.E., Swithenbank,
760	A., Castleton, M., Dewar, H., Mate, B.R., Shillinger, G.L., Schaefer, K.M., Benson,
761	S.R., Weise, M.J., Henry, R.W., and Costa, D.P. 2011. Tracking apex marine
762	predator movements in a dynamic ocean. Nature 475(7354): 86–90.
763	doi:10.1038/nature10082.
764	Burke, B.J., Liermann, M.C., Teel, D.J., and Anderson, J.J. 2013. Environmental and
765	geospatial factors drive juvenile Chinook salmon distribution during early ocean
766	migration. Can. J. Fish. Aq. Sci. 70: 1167–1177. doi:10.1139/cjfas-2012-0505.
767	Carlson, S.M., and Satterthwaite, W.H. 2011. Weakened portfolio effect in a collapsed
768	salmon population complex. Can. J. Fish. Aq. Sci. 68: 1579–1589.
769	doi:10.1139/f2011-084.
770	Carpenter, B., Gelman, A., Hoffman, M., Lee, D., Ben Goodrich, Betancourt, M.,
771	Brubaker, M.A., Guo, J., Li, P., and Riddell, A. 2017. Stan: A probabilistic
772	programming language. J. Stat. Softw. 76:1-32 doi:10.18637/jss.v076.i01
773	Chasco, B., I. Kaplan, A.C. Thomas, A. Acevedo-Gutiérrez, D Noren , M.J. Ford, M.B.
774	Hanson , Mr. J. Scordino , S. Jeffries, K.N. Marshall, A.O. Shelton, C. Matkin, B.
775	Burke, E.J. Ward. 2017. Competing tradeoffs between increasing marine mammal

ord.		
of reco	776	predation and fisheries harvest of Chinook salmon. Scientific Reports. 7: 15439.
ersion	777	doi:10.1038/s41598-017-14984-8.
ficial v	778	Coronado, C. and Hilborn, R. 1998. Spatial and temporal factors affecting survival in
15/18 inal of	779	coho and fall Chinook salmon in the Pacific Northwest. Bulletin of Marine Scier
on 06/ n the f	780	62:409–425.
ab Lib Ter froi	781	CTC 2015. 2014 Exploitation Rate Analysis and Model Calibration Volume One. Pa
nmal L nay dif	782	Salmon Commission Joint Chinook Technical Committee. Available at:
ne Man on. It r	783	http://www.psc.org/publications/technical-reports/technical-committee-
l Marir mpositi	784	reports/chinook/
Vationa age coi	785	Duffy, E.J. and Beauchamp D.A. 2011. Rapid growth in the early marine period
m by N g and p	786	improves the marine survival of Chinook salmon (Oncorhynchus tshawytscha) in
ress.co editing	787	Puget Sound, Washington. Can. J. Fish. Aquat. Sci. 68(2):232-240.
searchp to copy	788	Gelman, A., Lee, D., and Guo, J. 2015. Stan: A probabilistic programming language
v.nrcrei t prior	789	Bayesian inference and optimization. J. Educ. Behav. Stat. 40(5): 530-543.
m wwv iuscrip	790	doi:10.3102/1076998615606113.
led fro ed mar	791	Good, T.P., M.M. McClure, B.P. Sandford, K.A. Barnas, D.M. Marsh, B.A. Ryan, an
wnloac accept	792	Casillas. 2007 Quantifying the effect of Caspian tern predation on threatened and
Sci. Do t is the	793	endangered Pacific salmon in the Columbia River estuary. Endangered Species
vquat. S nuscrip	794	Research 3:11-21.
Fish. A	795	Greene, C.M., Hall, J.E., Guilbault, K.R., and Quinn, T.P. 2010. Improved viability of
Can. J. is Just	796	populations with diverse life-history portfolios. Biol. Lett. 6(3): 382–386.
nly. Th	797	doi:10.1098/rsb1.2009.0780.
personal use o	798	Griffiths, J.R., Schindler, D.E., Armstrong, J.B., Scheuerell, M.D., Whited, D.C., Cla
For		

598-017-14984-8. ilborn, R. 1998. Spatial and temporal factors affecting survival in ninook salmon in the Pacific Northwest. Bulletin of Marine Science ploitation Rate Analysis and Model Calibration Volume One. Pacific ssion Joint Chinook Technical Committee. Available at: org/publications/technical-reports/technical-committeechamp D.A. 2011. Rapid growth in the early marine period arine survival of Chinook salmon (Oncorhynchus tshawytscha) in ashington. Can. J. Fish. Aquat. Sci. 68(2):232-240. , and Guo, J. 2015. Stan: A probabilistic programming language for nce and optimization. J. Educ. Behav. Stat. 40(5): 530–543. 6998615606113. IcClure, B.P. Sandford, K.A. Barnas, D.M. Marsh, B.A. Ryan, and E. Quantifying the effect of Caspian tern predation on threatened and ific salmon in the Columbia River estuary. Endangered Species 1. J.E., Guilbault, K.R., and Quinn, T.P. 2010. Improved viability of n diverse life-history portfolios. Biol. Lett. 6(3): 382–386. .2009.0780. ndler, D.E., Armstrong, J.B., Scheuerell, M.D., Whited, D.C., Clark,

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/15/18 final of	802	
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Can. This Ju	819	
only.	820	
nal use	821	
r perso		
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nd.

- 00 Performance of salmon fishery portfolios across western North America. J. Appl.
- Ecol. 51(6): 1554–1563. doi:10.1111/1365-2664.12341.
- 802 Hankin, D.G. and Logan, E. 2010. A preliminary analysis of Chinook salmon coded-wire
- tag recovery data from Iron Gate, Trinity River and Cole Rivers Hatcheries, brood
- years 1978–2004. Prepared for The Hoopa Valley Tribal Council and the Arcata
- 805 Office, U.S. Fish and Wildlife Service. Available at:
- https://www.fws.gov/arcata/fisheries/reports/technical/IGHTRH.CWTanalysis2009%
- 807 20Hankin%20Rpt.pdf
- 808 Healey, M.C. 1991. Life history of Chinook salmon (Oncorhynchus tshawytscha). In:
- 809 Groot, C. and Margolis, L., eds. Pacific salmon life histories. Vancouver, BC. pp.
 810 311–394.
- Hilborn, R., Quinn, T.P., Schindler, D.E., and Rogers, D.E. 2003. Biocomplexity and
- fisheries sustainability. P. Natl. Acad. Sci. USA. 100(11): 6564–6568.
- doi:10.1073/pnas.1037274100.
- 814 Hoffman, M.D., and Gelman, A. 2014. The No-U-turn sampler: adaptively setting path
- lengths in Hamiltonian Monte Carlo. J. Mach. Learn. Res. 15: 1593–1623.
- Johnson, J.K. 1990. Regional overview of coded wire tagging of anadromous salmon and
- steelhead in northwest America. Am. Fish. Soc. Symp. 7: 782–816.
- 818 Kilduff, D.P., Botsford, L.W., and Teo, S.L.H. 2014. Spatial and temporal covariability
- in early ocean survival of Chinook salmon (*Oncorhynchus tshawytscha*) along the
- west coast of North America. ICES J. Mar. Sci. 71(7): 1671–1682.
 - doi:10.1093/icesjms/fsu031.

Can. J. Fish. Aquat. Sci. Downloaded from www.nrcresearchpress.com by National Marine Mammal Lab Lib on 06/15/18 For personal use only. This Just-IN manuscript is the accepted manuscript prior to copy editing and page composition. It may differ from the final official version of record.

822	Kilduff, D.P., Di Lorenzo, E., Botsford, L.W., and Teo, S.L.H. 2015. Changing central
823	Pacific El Niños reduce stability of North American salmon survival rates. P. Natl.
824	Acad. Sci. 112(35): 10962–10966. doi:10.1073/pnas.1503190112.
825	Koellner, T., and Schmitz, O.J. 2006. Biodiversity, ecosystem function, and investment
826	risk. BioScience 56(12): 977–985. doi:10.1641/0006-
827	3568(2006)56[977:BEFAIR]2.0.CO;2.
828	Lindley, S.T., Grimes, C.B., Mohr, M.S., Peterson, W., Stein, J., Anderson, J.T.,
829	Botsford, L.W., Bottom, D.L., Busack, C.A., Collier, T.K. et al. 2009. What caused
830	the Sacramento River fall Chinook stock collapse? NOAA Tech. Memo. NOAA-TM-
831	NMFS-SWFSC-447.
832	Markowitz, H. 1952. Portfolio selection. J. Financ. 7(1): 77-91. doi:10.1111/j.1540-
833	6261.1952.tb01525.x.
834	Martin, T.G., Chadès, I., Arcese, P., Marra, P.P., Possingham, H.P., and Norris, D.R.
835	2007. Optimal conservation of migratory species. PLoS ONE 2(8): e751.
836	doi:10.1371/journal.pone.0000751.
837	Monnahan, C.C., Thorson, J.T., and Branch, T.A. 2016. Faster estimation of Bayesian
838	models in ecology using Hamiltonian Monte Carlo. Meth. Ecol. Evol. 8: 339-348.
839	doi:10.1111/2041-210X.12681.
840	Moore, J.W., McClure, M., Rogers, L.A., and Schindler, D.E. 2010. Synchronization and
841	portfolio performance of threatened salmon. Conserv. Lett. 3(5): 340-348.
842	doi:10.1111/j.1755-263X.2010.00119.x.
843	Moran, P., Teel, D.J., Banks, M.A., Beacham, T.D., Bellinger, R.M., Blankenship, S.M.,

844 Candy, J.R., Garza, J.C., Hess, J.E., Narum, S.R., Seeb, L.W., Templin, W.D.,

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845 Wallace, C.G., and Smith, C.T. 2012. Divergent life-history races do not represent 846 Chinook salmon coast-wide: the importance of scale in Quaternary biogeography. 847 Can. J. Fish. Aq. Sci. 70: 415–435. doi:10.1139/cjfas-2012-0135. 848 Myers, J.M., Kope, R.G., Bryant, G.J., and Teel, D. 1998. Status review of Chinook 849 salmon from Washington, Idaho, Oregon, and California. NOAA Technical Memo 850 NMFS-NWFSC-35. 851 Naish, K.A., Taylor, J.E., III, Levin, P.S., Quinn, T.P., Winton, J.R., Huppert, D., and 852 Hilborn, R. 2007. An evaluation of the effects of conservation and fishery 853 enhancement hatcheries on wild populations of salmon. Adv. Mar. Biol. 53: 61–194. 854 doi:10.1016/S0065-2881(07)53002-6. 855 Nandor, G.F., Longwill, J.R., and Webb, D.L. 2010. Overview of the coded wire tag 856 program in the greater Pacific region of North America. In: Wolf, K.S. and O'Neal, 857 J.S., eds., PNAMP special publication: tagging, telemetry, and marking measures for 858 monitoring fish populations. Pacific Northwest Aquatic Monitoring Partnership 859 Special Publication 2010-002. pp. 5-46. 860 Neal, R.M. 2011. MCMC using Hamiltonian dynamics. In: Brooks, S., Gelman, A., 861 Jones, G.L., and Meng, X.-L. eds. Handbook of Markov Chain Monte Carlo. 862 Handbook of Markov Chain Monte Carlo. pp. 113–162. CRC Press. 863 Newman, K.B. 1998. State-space modeling of animal movement and mortality with 864 application to salmon. Biometrics 54(4): 1290. doi:10.2307/2533659. 865 Newman, K.B. 2000. Hierarchic modeling of salmon harvest and migration. J. Agr. Biol. 866 Envir. St. 5(4): 430-455. doi:10.2307/1400659. 867 Norris, J.G., Hyun, S.Y., and Anderson, J.J. 2000. The ocean distribution of the

Page 39 of 49

868	Columbia River upriver bright fall chinook salmon stocks. North Pacific
869	Anadromous Fish Commission Bulletin, 2: 221–232.
870	PFMC (Pacific Fishery Management Council). 2016a. Review of 2015 Ocean Salmon
871	Fisheries Stock Assessment and Fishery Evaluation Document for the Pacific Coast
872	Salmon Fishery Management Plan. Pacific Fishery Management Council, 7700 NE
873	Ambassador Place, Suite 101, Portland, OR 97220-1384, USA.
874	PFMC (Pacific Fishery Management Council). 2016b. Pacific Coast salmon fishery
875	management plan for commercial and recreational salmon fisheries off the coasts of
876	Washington, Oregon, and California as revised through amendment 19. Pacific
877	Fishery Management Council, 7700 NE Ambassador Place, Suite 101, Portland,
878	Oregon 97220-1384.
879	Pacific Salmon Commission. 2016. Joint Chinook Technical Committee annual report of
880	catch and escapement for 2015. Report TCCHINOOK (16)-3. Pacific Salmon
881	Commission, 600 - 1155 Robson Street, Vancouver, B.C., Canada V6E 1B5
882	Quinn, T.P. 2005. The Behavior and Ecology of Pacific Salmon and Trout. University of
883	Washington Press, Canada.
884	R Core Team. 2016. R: A language and environment for statistical computing. R
885	Foundation for Statistical Computing, Vienna, Austria. URL https://www.R-
886	project.org/.
887	Ruckelshaus, M.H., Levin, P., Johnson, J.B., and Kareiva, P.M. 2003. The Pacific salmon
888	wars: what science brings to the challenge of recovering species Annu. Rev. Ecol.
889	Syst. 33:665-706. doi:10.1146/annurev.ecolsys.33.010802.150504.
890	Rue, H., Martino, S., Chopin, N. 2009. Approximate Bayesian inference for latent

891 Gaussian models using integrated nested Laplace approximations (with discussion).

B92 J. Roy. Stat. Soc. B. 71:319-392.

893 Satterthwaite, W.H., and Carlson, S.M. 2015. Weakening portfolio effect strength in a

hatchery-supplemented Chinook salmon population complex. Can. J. Fish. Aq. Sci.

895 72: 1860-1875. doi:10.1139/cjfas-2015-0169.

896 Satterthwaite, W.H., Ciancio, J., Crandall, E., Palmer-Zwahlen, M.L., Grover, A.M.,

897 O'Farrell, M.R., Anderson, E.C., Mohr, M.S., and Garza, J.C. 2015. Stock

898 composition and ocean spatial distribution inference from California recreational

Chinook salmon fisheries using genetic stock identification. Fish. Res. 170: 166–178.
doi:10.1016/j.fishres.2015.06.001.

Satterthwaite, W.H., Mohr, M.S., O'Farrell, M.R., and Wells, B.K. 2013. A comparison
of temporal patterns in the ocean spatial distribution of California's Central Valley
Chinook salmon runs. Can J Fish Aq Sci 70(4): 574–584. doi:10.1139/cjfas-20120395.

905 Satterthwaite, W.H., Mohr, M.S., O'Farrell, M.R., Anderson, E.C., Banks, M.A., Bates,

906 S.J., Bellinger, M.R., Borgerson, L.A., Crandall, E.D., Garza, J.C., Kormos, B.J.,

907 Lawson, P.W., and Palmer-Zwahlen, M.L. 2014. Use of genetic stock identification

data for comparison of the ocean spatial distribution, size at age, and fishery

909 exposure of an untagged stock and its indicator: California Coastal versus Klamath

910 River Chinook salmon. Trans. Am. Fish. Soc. 143: 117–133.

911 doi:10.1080/00028487.2013.837096.

Schindler, D.E., Armstrong, J.B., and Reed, T.E. 2015. The portfolio concept in ecology
and evolution. Front. Ecol. Env. 13: 257–263. doi:10.1890/140275.

ord.		
ofrec	914	Schindler, D.E., Armstrong, J.B., Bentley, K.T., Jankowski, K., Lisi, P.J., and Payne,
rersion	915	L.X. 2013. Riding the crimson tide: mobile terrestrial consumers track phenological
ficial v	916	variation in spawning of an anadromous fish. Biol. Lett. 9:20130048.
/15/18 final of	917	doi:10.1098/rsbl.2013.0048.
on 06/ m the	918	Schindler, D.E., Hilborn, R., Chasco, B., Boatright, C.P., Quinn, T.P., Rogers, L.A., and
ab Lib ffer frc	919	Webster, M.S. 2010. Population diversity and the portfolio effect in an exploited
mmal I may di	920	species. Nature 465: 609-612. doi:10.1038/nature09060.
ine Ma tion. It	921	Sharma, R., Vélez Espino, L.A., Wertheimer, A.C., Mantua N., and Francis, R.C. 2013.
al Mari mposi	922	Relating spatial and temporal scales of climate and ocean variability to survival of
Nation page cc	923	Pacific Northwest Chinook salmon (Oncorhynchus tshawytscha). Fisheries
om by ig and	924	Oceanography 22: 14–31. doi:10.1111/fog.12001.
press.c y editir	925	Stan Development Team. 2016. RStan: the R interface to Stan, Version 2.10.1. http://mc-
to cop	926	stan.org.
w.nrcre ot prior	927	Teel, D.J., Burke, B.J., Kuligowski, D.R., Morgan, C.A., and Van Doornik, D.M. 2015.
inuscrij	928	Genetic identification of Chinook Salmon: stock-specific distributions of juveniles
ded frc ted ma	929	along the Washington and Oregon coasts. Mar. Coast. Fish. 7:274–300
ownloa e accep	930	doi:10.1080/19425120.2015.1045961.
Sci. Do pt is th	931	Thorson, J.T., Scheuerell, M.D., Buhle, E.R., and Copeland, T. 2014. Spatial variation
Aquat. inuscri	932	buffers temporal fluctuations in early juvenile survival for an endangered Pacific
Fish.	933	salmon. J. Anim. Ecol. 83(1): 157-167. doi:10.1111/1365-2656.12117.
Can. J his Just	934	Trudel, M., Fisher, J., Orsi, J.A., Morris, J.F.T., Thiess, M.E., Sweeting, R.M., Hinton,
nly. T	935	S., Fergusson, E.A., and Welch, D.W. 2009. Distribution and migration of juvenile
al use c	936	Chinook salmon derived from coded wire tag recoveries along the continental shelf
erson		

- of western North America. Trans. Am. Fish. Soc. 138(6): 1369–1391.
- 938 doi:10.1577/T08-181.1.
- 939 Tucker, S., Trudel, M., Welch, D.W., Candy, J.R., Morris, J.F.T., Thiess, M.E., Wallace,
- 940 C., and Beacham, T.D. 2011. Life history and seasonal stock-specific ocean
- 941 migration of juvenile Chinook Salmon. Trans. Am. Fish. Soc. 140(4): 1101–1119.
- 942 doi:10.1080/00028487.2011.607035.
- 943 Waples, R.S., Teel, D.J., Myers, J.M., and Marshall, A.R. 2004. Life-history divergence
- 944 in Chinook salmon: historic contingency and parallel evolution. Evolution 58(2):
- 945 386–403. doi:10.1111/j.0014-3820.2004.tb01654.x.
- 946 WDFW (Washinton Department of Fisheries). 1979. Washington State sport catch report.
- 947 Available at: http://wdfw.wa.gov/fishing/harvest/
- 948 Weitkamp, L.A. 2010. Marine distributions of Chinook salmon from the west coast of
- 949 North America Determined by coded wire tag recoveries. Trans. Am. Fish. Soc.
- 950 139(1): 147–170. doi:10.1577/T08-225.1.
- 951 Winans, G.A., Viele, D., Grover, A., Palmer-Zwahlen, M., Teel, D., and Van Doornik, D.
- 952 2001. An update of genetic stock identification of Chinook salmon in the Pacific
- 953 Northwest: test fisheries in California. Rev. Fish. Sci. 9(4): 213–237.
- 954 doi:10.1080/20016491101753.
- 955

Figure 1: Map of study area, hatchery locations (black dots), and 17 coastal regions used in the study. Locator map (left) attribution: Esri, DeLorme, GEBCO, NOAA NGDC, and other contributors. Main map attribution: Esri, NOAA NGDC, NOAA GSHHG, and other contributors.

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964 Figure 2: Example raw CPUE data for two releases. Left panel: Observed CPUE (fish per 965 boat-day) from commercial troll fisheries for fall Chinook salmon released from Coleman 966 National Fish Hatchery (SFB region) in 1980 (N=393,932 at release). Black indicates 967 region-season combinations with commercial troll fisheries but zero observed catches. 968 Grey indicates no commercial troll fishery occurred region-season combination. Right 969 panel: Observed CPUE from commercial troll fisheries for fall Chinook released from 970 Lyons Ferry (UPCOL region) released in 1984 (N=234,985 at release). Note that the 971 color ramp differs between panels.





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Figure 3: Estimated proportional spatial distribution by season of fall Chinook salmon originating from 11 different regions ($\theta_{l,r,s}$). Each row represents the proportion of fish from a region present in each ocean region (rows sum to one). Posterior means are shown.



985 Figure 4: Survivorship trends for juvenile Chinook salmon from release to August 1, 986 brood year +2 for different origin regions between 1978 and 1990. Raw survivorship 987 scores have been converted to z-scores (subtract the among-year mean, divide by the 988 standard deviation) to make trends in survival comparable among regions. Panels show 989 weighted mean for Chinook from northern regions (a), the Columbia basin (b), and 990 southern regions (c; see also Fig. 1). Individual points show mean estimates for 991 individual release groups. Due to low numbers of release groups in Oregon regions 992 (NOR, COR, SOR), we combined Oregon releases into a single "OR" region.

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Figure 5: Distribution and abundance of fall Chinook salmon age 3 and older in the ocean. We show proportional contribution of age 3+ fish to each ocean region (left panels) and total abundance (right panels) at the beginning of spring (*a*), summer (*b*), and fall (*c*) seasons. Results arise from simulations assuming median fishing mortality for each area and season (see Fig. S1.9) and a single juvenile mortality rate shared across all regions.



1008 Figure 6. Summer distribution and abundance of fall Chinook salmon from four 1009 representative regions and two age groups under the base scenario. Areas SFB (a, f), 1010 COL(b,g), WAC (c,h), SGEO (d,i), and aggregate abundance across all stocks (e,j)



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Figure 7: Comparison of two simulation scenarios. (a,b) Proportional contribution of age 3+ fish to each ocean region (left panels) and total abundance (right panels) at the beginning of summer under the base scenario (a) and under reduced hatchery production from the PUSO region (b). Panel (c) shows the proportional change in total abundance from base to PUSO hatchery scenarios for age 3+ Chinook salmon.



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